

ii. Increase the flow rate to 2 to 4 milliliters per minute by applying air pressure to the column. A glycerol manostat adjusted to 30 inches and attached between an air supply and column provides adequate pressure.

b. Wash the resin with 10 milliliters of eluting reagent A. Discard eluate.

2. Pass eluate A from procedure step V–D4 through the column. Collect in a 250-milliliter beaker.

3. Pass 50 milliliters of specially denatured alcohol 3A through the column. Combine with the eluate of procedure step V–E2.

F. Reduction. 1. Place the eluate A fraction from procedure step V–E3 on a hot water bath (90° C.) and evaporate with a stream of air until 5 to 10 milliliters remain. Do not overheat the sample or allow the sample to go to dryness.

2. Transfer to centrifuge tube and rinse beaker three times with 3 milliliters of specially denatured alcohol 3A.

3. Evaporate on a hot water bath (90° C.) under a stream of air until alcohol has evaporated. Do not overheat the sample or allow the sample to go to dryness.

4. Remove the tube from the water bath and immediately add 5.0 milliliters of water.

5. While mixing, add 2 drops of titanium chloride and 4 drops of 10*N* sodium hydroxide. Continue mixing until greyish color disappears.

a. Mix on Vortex Jr. mixer, or equivalent, regulated with autotransformer.

b. Precipitate of insoluble tissue substances and white titanium salts is present after reduction is complete.

6. Dilute to 50 milliliters with specially denatured alcohol 3A and mix.

7. Centrifuge for 5 minutes at 2,000 r.p.m.

G. Cation-exchange chromatography—No. 2. 1. Prepare resin column by procedure step V–E.

2. Pass the centrifugate of procedure step V–F7 through column. Use three rinses of specially denatured alcohol 3A, each 5 milliliters, to aid in transferring of sample.

3. Pass 50 milliliters of specially denatured alcohol 3A through the column.

4. Pass 50 milliliters of deionized water through the column.

5. Elute arylamine residue from the resin with 40 to 43 milliliters of 4*N* HCl into a 50-milliliter volumetric flask (actinic ware) for 3,5-DNBA analysis. Avoid direct sunlight. The arylamine has been found to be photosensitive.

H. Color development and measurement. 1. Cool to 0° C.–5° C. by placing in a freezer or ice bath.

2. Perform the Bratton-Marshall reaction in subdued light as follows:

a. Add 1 milliliter of sodium nitrite reagent, mix, and allow to stand for 1 minute.

b. Add 1 milliliter of ammonium sulfamate reagent, mix, and allow to stand for 1 minute.

c. Add 1 milliliter of coupling reagent, mix, and allow to stand for 10 minutes.

d. Dilute to volume with 4*N* HCl.

3. Perform colorimetric measurement at 530 millimicrons as follows:

a. Fill two matched 100-millimeter cells with 4*N* HCl and place into instrument.

b. Adjust dark current.

c. Adjust to zero absorbance.

d. Replace acid in cell of sample side of compartment with sample to be measured.

e. Record absorbance observed.

I. Calculations. Determine parts per billion (observed) from the standard curve.

§ 556.225 Doramectin.

A tolerance of 0.1 part per million (ppm) is established for parent doramectin (marker residue) in liver (target tissue) of cattle and 0.16 ppm in liver of swine.

[62 FR 62243, Nov. 21, 1997]

§ 556.227 Eprinomectin.

Tolerances are established for residues of eprinomectin B1a (marker residue) in milk of 12 parts per billion and in liver (target tissue) of 4.8 parts per million.

[62 FR 33998, June 24, 1997]

§ 556.228 Enrofloxacin.

A tolerance of 0.3 part per million is established for residues of enrofloxacin (marker residue) in muscle (target tissue) of chickens and turkeys.

[61 FR 56893, Nov. 5, 1996]

§ 556.230 Erythromycin.

Tolerances for residues of erythromycin in food are established as follows:

(a) 0.1 part per million in uncooked edible tissues of beef cattle and swine.

(b) Zero in milk.

(c) 0.025 part per million in uncooked eggs.

(d) 0.125 part per million (negligible residue) in uncooked edible tissues of chickens and turkeys.

[40 FR 13942, Mar. 27, 1975, as amended at 58 FR 43795, Aug. 18, 1993]

§ 556.240 Estradiol and related esters.

No residues of estradiol, resulting from the use of estradiol or any of the related esters, are permitted in excess of the following increments above the

concentrations of estradiol naturally present in untreated animals:

(a) In uncooked edible tissues of heifers, steers, and calves:

- (1) 120 parts per trillion for muscle.
- (2) 480 parts per trillion for fat.
- (3) 360 parts per trillion for kidney.
- (4) 240 parts per trillion for liver.

(b) In uncooked edible tissues of lambs:

- (1) 120 parts per trillion for muscle.
- (2) 600 parts per trillion for fat, kidney, and liver.

[49 FR 13873, Apr. 9, 1984, as amended at 56 FR 67175, Dec. 30, 1991]

§ 556.260 Ethopabate.

Tolerance for residues of ethopabate converted to metaphenethidine are established in the edible tissues of chickens as follows:

(a) 1.5 parts per million in uncooked liver and kidney.

(b) 0.5 part per million in uncooked muscle.

§ 556.270 Ethylenediamine.

A tolerance of zero is established for residues of ethylenediamine in milk.

§ 556.273 Famphur.

Tolerances are established for residues of famphur including its oxygen analog in or on meat, fat, or meat by-products of cattle at 0.1 part per million.

[62 FR 55161, Oct. 23, 1997]

§ 556.275 Fenbendazole.

(a) *Cattle and goats.* A tolerance¹ of 0.8 part per million is established for parent fenbendazole (the marker residue) in the liver of cattle and goats.

(b) *Swine.* A tolerance¹ for marker residues of fenbendazole in swine is not needed.

(c) *Cattle milk.* A safe concentration of 1.67 parts per million is established for total fenbendazole residues. A tolerance of 0.6 part per million is estab-

¹As used in this section: "tolerance" refers to a concentration of a marker residue in the target tissue selected to monitor for total residues of the drug in the target animal.

lished based on the fenbendazole sulfide metabolite (marker residue).

[59 FR 26943, May 25, 1994, as amended at 61 FR 29478, June 11, 1996]

§ 556.277 Fenprostalene.

A tolerance for marker residue of fenprostalene in cattle is not needed. The safe concentrations for the total residues of fenprostalene in the uncooked edible tissues of cattle are 10 parts per billion in muscle, 20 parts per billion in liver, 30 parts per billion in kidney, 40 parts per billion in fat, and 100 parts per billion in the injection site. As used in this section "tolerance" refers to a concentration of a marker residue in the target tissue selected to monitor for total residues of the drug in the target animal, and "safe concentrations" refer to the concentrations of total residues considered safe in edible tissues.

[49 FR 26716, June 29, 1984]

§ 556.283 Florfenicol.

The safe concentrations for total florfenicol-related residues in cattle are 2.0 parts per million (ppm) in muscle, 6.0 ppm in liver, and 12.0 ppm in kidney and fat. A tolerance of 3.7 ppm for the marker residue, florfenicol amine, has been established in cattle liver.

[61 FR 42383, Aug. 15, 1996]

§ 556.290 Furazolidone.

A tolerance of zero is established for residues of furazolidone in the uncooked edible tissues of swine.

§ 556.300 Gentamicin sulfate.

(a) A tolerance of 0.1 part per million is established for negligible residues of gentamicin sulfate in the uncooked edible tissues of chickens and turkeys.

(b) Tolerances are established for total residues of gentamicin in edible tissues of swine as follows: 0.1 part per million in muscle, 0.3 part per million in liver, and 0.4 part per million in fat and kidney. A microbiological determinative procedure and an HPLC confirmatory procedure for gentamicin have been developed to assay gentamicin in kidney at 0.4 ppm. Since residues of gentamicin as the parent compound and total residues are equal,